



PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of

Xiao and Gedrich

Serial No. 09/879,792

Filing Date: June 13, 2001

)
) Group Art Unit: 1652
)
) Examiner: D. Ramirez
)
) Docket No. 002973.00035
)

RECEIVED
JUN 11 2003
TECH CENTER 1600/2900

For: **REGULATION OF HUMAN TRANSMEMBRANE SERINE PROTEASE**

DECLARATION UNDER RULE 131

We, Yonghong Xiao and Richard Gedrich, hereby declare:

1. We are the named inventors of the subject matter claimed in the application referenced above.
2. Prior to March 19, 2001, we conceived of and reduced to practice molecules comprising the polynucleotide sequence disclosed as SEQ ID NO:11 and the amino acid sequence disclosed as SEQ ID NO:12 in the above-referenced application.
3. The nucleotide sequence of a contig identified as SEQ ID NO:11 in serial number 60/211,224 was extended by amplification and alignments with mouse EST and human genomic sequences. The nucleotide sequence of an extended contig containing a portion of the sequence of SEQ ID NO:11 in serial number 09/879,792 was identified. The nucleotide sequence of the extended contig is shown on the attached copy of page 3 of laboratory notebook RB55202 and identified as "147.33 contig" (Exhibit A).
4. The Basic Local Alignment Search Tool (BLAST) algorithm was used to extend the 5' end sequence of the 147.33 contig. The BLAST algorithm aligned human expressed sequence tags (ESTs) having sequence similarity with the 5' sequence of the 147.33 contig. The results of the BLAST searches were recorded on pages 20-23 of laboratory notebook RB55202. Copies are provided as Exhibit B.
5. An EST, BE280394, was found to overlap the 5' sequence of the 147.33 contig. The BE280394 EST sequence was recorded on page 24 of laboratory notebook RB55202. A copy

of page 24 is provided as Exhibit C. The alignment of EST BE280394 with the 5' end of the contig is shown in second column of page 20, the first column of page 21, and the first lines of the second column of page 21 (Exhibit B).

6. To confirm that the 147.33 contig extended at its 5' end with the sequence of EST BE280394 included the sequence of a full-length gene, the extended 147.33 contig was translated by computer into an amino acid sequence. The translation of the extended contig in all possible open reading frames associated with the extended sequence was recorded on pages 27 and 28 of laboratory notebook RB55202. Copies are provided as Exhibit D. The extended contig was confirmed to contain a full-length gene because its translation into an amino acid sequence identified a start (methionine) residue at its amino terminal end and the start codon is in frame with a predicted amino acid sequence encoded by the 147.33 contig. The translated sequence identified as the one encoding the protein shown in SEQ ID NO:12 is underlined and begins "MERDSH." The hand-written notes at the top of page 29 of laboratory notebook RB55202 identify the extended contig sequence as the sequence of the "147" full-length gene. A copy of page 29 is included in Exhibit D.

7. Primers capable of amplifying the 147 full-length open reading frame were ordered based on the nucleotide sequence that encodes the translated protein. The oligonucleotide sequences ordered were recorded on page 30 of laboratory notebook RB55202, a copy of which is provided as Exhibit E.

8. Human placenta cDNA was amplified using these primers to obtain the molecule in human genomic DNA that contains the 147 full-length open reading frame. The primers used to perform the amplification and the conditions of the amplification reaction were recorded on page 40 of laboratory notebook RB55202. A copy of page 40 is provided as Exhibit F.

9. The amplification product obtained from this reaction was the predicted size for the full-length open reading frame. An agarose gel showing the amplification product obtained using the primers is shown in a photograph on page 41 of laboratory notebook RB55202. A copy of page 41 is provided as Exhibit G.

10. The amplification product was extracted from the gel using the "QIAquick Gel Extraction Kit Protocol" and cloned into a vector sold by Invitrogen and referred to the "TA vector." The protocol used to clone the amplification product (full-length open reading frame)

into the vector was recorded on pages 41 and 42 of lab notebook RB55202. Copies of pages 41 and 42 are provided as Exhibits G and H, respectively.

11. The vector containing the amplification product was transformed into *E. coli* TOP10 cells, as described on page 42 of laboratory notebook RB55202 (Exhibit H).

12. The vector was extracted from the transformed *E. coli* and digested with restriction enzyme *EcoRI* to confirm the presence of the amplification product in the vector. The protocol "QIAprep Spin Miniprep Kit Protocol," which was used to obtain the vector DNA from *E. coli*, was recorded on page 1 of laboratory notebook RB55846. A copy of page 1 is included in Exhibit I. The conditions under which the vector DNA was digested and a picture of the agarose gel showing the results of restriction enzyme digestion are provided on page 7 of laboratory notebook RB55846. A copy of this page is included in Exhibit I. The agarose gel on the laboratory notebook page confirms the presence of the 147 full-length open reading frame in each vector but for one.

13. Vectors containing the amplification product were sequenced. The composition of the tubes containing the vector and primers used in the sequencing reaction were recorded on page 8 of laboratory notebook RB55846. A copy of page 8 is provided as Exhibit J.

14. The nucleotide sequence obtained for amplification product cloned into the vector was recorded on pages 15-17 of laboratory notebook RB55846. Copies of pages 15-17 are provided as Exhibit K. The recorded nucleotide sequence is identical to the sequence identified as SEQ ID NO:11 in application serial number 09/879,792.

15. The nucleotide sequence of the amplification product was translated into an amino acid sequence. This translated amino acid sequence was recorded on pages 17-18 of laboratory notebook RB55846. Copies of pages 17-18 are provided as Exhibit L. The recorded amino acid sequence is identical to the sequence identified as SEQ ID NO:12 in application serial number 09/879,792.


16. The dates on each of the attached Exhibits have been redacted. All of the work described in paragraphs 3 through 15 was performed in the United States and completed before March 19, 2001.

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these were made with the knowledge that false statements made willfully are punishable by fine, imprisonment, or both a fine and imprisonment under Section 1001 of Title 18 of the United States; and further that false statements made willfully may jeopardize the validity of any patent issuing on an application in which the false statements were made.

Date

Yonghong Xiao

6/5/03
Date

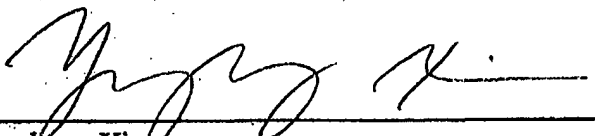


Richard Gedrich

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these were made with the knowledge that false statements made willfully are punishable by fine, imprisonment, or both a fine and imprisonment under Section 1001 of Title 18 of the United States; and further that false statements made willfully may jeopardize the validity of any patent issuing on an application in which the false statements were made.

6/9/03

Date


Yonghong Xiao~~6/9/03~~

Date


Richard Gedrich

BAYER CORPORATION

Note Book 3
RB55202

SUBJECT

147.33.contig2 Sequence (Nucleotide)

EXHIBIT

A

147.33.contig2
(Strand)

1 GAATTCGCTT TTTCTTTCAG AATGCAATTC CAGCAAGAAC
41 ACCTTCAGCT GGAGCATCTC CAGCCGAGGT ATCTCCAGCT
61 GGAGCACTTC CAGCCGAGGT ATCTCCAGCT CAGGATATTC
121 CAGCCGAGGT ATCTCCAGCT CAGGATATTC CAGGATATTC
161 ATCTCCAGCT CAGGATATTC CAGGATATTC ACCTCCAGCT
201 CAGGATATTC CAGGATATTC ATCTCCAGCT CAGGATATTC
241 CAGGATATTC ATCTCCAGCT CAGGATATTC ATCTCCAGCT
281 CTGATCCGCT AGGTCATCAT CCGCCAGGTC AGCTTCGGTG
321 ACAAGCTGCT CAACGAGAGT GTACCTTCCT AGAGCAACAC
361 CAGTGGGGCT TGTACCATTC CAGTTCATTC CTGCGAGTC
401 AGCAGCAGCA ACCGAGCAGC CAGGAGCAGC CAGGAGCAGC
441 AGCTTCGCTC ACTTCAGCTC CCGGGAGGTC CAGGAGCAGC
481 TACCGCTCAT TCGGTCGCTC CTCTTCCTCA TTGCGCTGCT
521 GCTTTCGCTC ATCATCTCTC TCGAGTTCCT CAGGAGCAGC
561 ACAGGGATCA GGTCAAGGCA CCGAGGAGTC AGCTTCGCTC
601 AGCAGCTGCT CCGGTCGCTC CTCTTCCTCA TTGCGCTGCT
641 GAAGATGAGC CCGGTCGCTC CTCTTCCTCA TTGCGCTGCT
681 AAGTCTCTCT TCAAAATCTA CTCTTCGCTC TCGCATCATC
721 GGTCTTCCTC CTCTTCGAGC AACTGGAATG AGCTTCATCT
761 ACAGAGAGCT TCGCAGCAGC TCGGTCGCTC ATCTTCCTCA
801 CCGACACAGC AGGTCGCTC CAGGAGATTC CCGACAGCT
841 TCTCAATCTT GAGATCAAGC TCGCAGCAGC AGCAGAGCT
881 CCGAGAGCTT GAATGCTCTT CCGAGGCTCA TATCTTCCTC
921 CAGTTCGCTC ACTTCGCTC CAGGAGATTC ACCTTCGCTC
961 TCGTGGGAGG GCGCTTCGCT TCGCATGAGC AGTTCGCTC
1001 GCAAGTGAAT CTGCTTCGCT CCGCAGCAGC CATCTTCGCA
1041 GCGACGCTCA TTGAGCCCCA GTGGGTCGCT ACTTCGCTC
1081 ACTGCTCTCT CCGCAGCAGC GAGAGGTCCT TCGGTCGCTC
1121 CAGGCTCTAC CCGGAGCAGC GAGAGGTCCT CAGGTCGCTC
1161 CAGGAGGCTT CCGATTCGCA CATCATCATC AACAGCAAT
1201 ACAGGAGTGA GAGAGGAGC TATGAGATTC CCGTTCGCTC
1241 GGTTCGCAAG CCGGTCGCTC TCGGTCGCTC CATTCGCTC
1281 GGTTCGCTC CCGATTCGCA ACAGAGCTTT AGCTTCGCTC
1321 AGAGCTCTCT GATCAGAGC TTGCGGAGC CCGGAGGAGC
1361 AGATCAGAG ACATTCGCTT TCGTTCGCTC GGTTCAGGTC
1401 AATCTCATCG ACTTCAGGAA ATGCAATGAC TACTTCGCTC
1441 ATGAGAGTGA CTTTACGCTC AGGATCATCT GTGCTGGGCA
1481 CTTTCGCTC GCGAGAGACT CCGGTCGCTC AGAGAGGCTC
1521 CCGGCTCTCT TCGTTCGCA GAGAGGCTC TCGTTCGCTC
1561 CAGGTCGCTC CAGGTCGCTC CCGGTCGCTC CCGAGAGAA
1601 CAAAGCTGCT CCGTTCAGCA AAGTTCAGCA ACTTCGCTC
1641 TCGATTTACA CCGATTCGCA GAGGAGGTC CCGATTCAGAA
1681 AATCTTAACC AGCTTCGCTC CCGTTCGCA CAGGAGGCTC
1721 TCTTCGCTC AAGGGGCAAT TC

LifeTools Reverse FASTA Sequences



Reverse
FASTA Sequences

Project ID: 000159147 Project Name: TM Series Proteome
Created By: Gerdien, Richard Created By: Gerdien, Richard
Modified By: Gerdien, Richard Modified By: Gerdien, Richard
Current User: Gerdien, Richard (WHITE)
Today's Date: Num Seqs: 26

Reverse(147.33.contig2):
GAATTCGCTT TTTCTTTCAG AATGCAATTC CAGCAAGAAC
GGAGCATCTC CAGCCGAGGT ATCTCCAGCT CAGGATATTC
CAGCCGAGGT ATCTCCAGCT CAGGATATTC CAGGATATTC
ATCTCCAGCT CAGGATATTC CAGGATATTC ACCTCCAGCT
CAGGATATTC CAGGATATTC ATCTCCAGCT CAGGATATTC
ATCTCCAGCT CAGGATATTC ATCTCCAGCT CAGGATATTC
CTGATCCGCT AGGTCATCAT CCGCCAGGTC AGCTTCGGTG
ACAAGCTGCT CAACGAGAGT GTACCTTCCT AGAGCAACAC
CAGTGGGGCT TGTACCATTC CAGTTCATTC CTGCGAGTC
AGCAGCAGCA ACCGAGCAGC CAGGAGCAGC CAGGAGCAGC
AGCTTCGCTC ACTTCAGCTC CCGGGAGGTC CAGGAGCAGC
TACCGCTCAT TCGGTCGCTC CTCTTCCTCA TTGCGCTGCT
GCTTTCGCTC ATCATCTCTC TCGAGTTCCT CAGGAGCAGC
ACAGGGATCA GGTCAAGGCA CCGAGGAGTC AGCTTCGCTC
AGCAGCTGCT CCGGTCGCTC CTCTTCCTCA TTGCGCTGCT
GAAGATGAGC CCGGTCGCTC CTCTTCCTCA TTGCGCTGCT
AAGTCTCTCT TCAAAATCTA CTCTTCGCTC TCGCATCATC
GGTCTTCCTC CTCTTCGAGC AACTGGAATG AGCTTCATCT
ACAGAGAGCT TCGCAGCAGC TCGGTCGCTC ATCTTCCTCA
CCGACACAGC AGGTCGCTC CAGGAGATTC CCGACAGCT
TCTCAATCTT GAGATCAAGC TCGCAGCAGC AGCAGAGCT
CCGAGAGCTT GAATGCTCTT CCGAGGCTCA TATCTTCCTC
CAGTTCGCTC ACTTCGCTC CAGGAGATTC ACCTTCGCTC
TCGTGGGAGG GCGCTTCGCT TCGCATGAGC AGTTCGCTC
GCAAGTGAAT CTGCTTCGCT CCGCAGCAGC CATCTTCGCA
GCGACGCTCA TTGAGCCCCA GTGGGTCGCT ACTTCGCTC
ACTGCTCTCT CCGCAGCAGC GAGAGGTCCT TCGGTCGCTC
CAGGCTCTAC CCGGAGCAGC GAGAGGTCCT CAGGTCGCTC
CAGGAGGCTT CCGATTCGCA CATCATCATC AACAGCAAT
ACAGGAGTGA GAGAGGAGC TATGAGATTC CCGTTCGCTC
GGTTCGCAAG CCGGTCGCTC TCGGTCGCTC CATTCGCTC
GGTTCGCTC CCGATTCGCA ACAGAGCTTT AGCTTCGCTC
AGAGCTCTCT GATCAGAGC TTGCGGAGC CCGGAGGAGC
AGATCAGAG ACATTCGCTT TCGTTCGCTC GGTTCAGGTC
AATCTCATCG ACTTCAGGAA ATGCAATGAC TACTTCGCTC
ATGAGAGTGA CTTTACGCTC AGGATCATCT GTGCTGGGCA
CTTTCGCTC GCGAGAGACT CCGGTCGCTC AGAGAGGCTC
CGGCTCTCT TCGTTCGCA GAGAGGCTC TCGTTCGCTC
CAGGTCGCTC CAGGTCGCTC CCGGTCGCTC CCGAGAGAA
CAAAGCTGCT CCGTTCAGCA AAGTTCAGCA ACTTCGCTC
TCGATTTACA CCGATTCGCA GAGGAGGTC CCGATTCAGAA
AATCTTAACC AGCTTCGCTC CCGTTCGCA CAGGAGGCTC
TCTTCGCTC AAGGGGCAAT TC

Requested by: Gerdien, Richard on Tue

INCYTE PHARMACEUTICALS, INC.

SIGNED BY

[Signature]

DATE

WITNESSED AND UNDERST

D BY

Elizabeth A. Sullivan

DATE

CROSS REFERENCES:

BAYER CORPORATION

RB55202 21

SUBJECT

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DATE

RB55202 23

doi:10.1371/journal.pone.0198323.g004 <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC6121463/> . DOI: 10.1371/journal.pone.0198323.g004

```
Matrix: blasr matrix: -3
Cap Penalties: Distance: 5, Extension: 3
Number of Hits: 26: 710245
Number of Sequences: 2774091
Number of Extensions: 710245
Number of successful extensions: 194161
Number of sequences better than 10.0: 28
Length of query: 787
Database: 1,181,000,365
effective length: 20
effective length of query: 767
effective length of database: 1,127,518.54
effective search space: 8600726015
effective search space used: 86400726015
T: 0
K: 0
X1: 5 (11.3 bits)
X2: 30 (19.4 bits)
X3: 22 (24.3 bits)
X4: 19 (38.2 bits)
```

- I will Transfer it to see
if it contains an in Form
ATG.

WITNESSED AND UNDERSTOOD BY Charles H. C. Hollins Notary

DATE _____

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EXHIBIT

D

LifeTools

DATE: 10/10/2018 10:10:10 AM PAGE: 10 OF 10

Page 5-1	Issue 1	INTRA	Classified	Sec. Agency	Group	Translation
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[illegible]

✓ COOLERT: 47, NE200J4:2

Sequence: coolERT147:RE280394:2

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-1 K P G L F P H E T M E R S H O N A S
-2 E L E D S R Q R Y W R O T A C G M H L
-3 A M R T F L R H O R G A T G C P R E C I
A A C C G A C C T T T C C T G C G C A C C A C C A C C G A A G A C C A C T
T T C G A C C T C T G A A G A G G C T C T G T A C C T C C A C C C G C C T A C T A G
-3 C P F S K O B S W M S I S L P A S A E
-1 A O L V R G R L S W F S P C O R S H A E
-2 L R S S C K O L H L F V A V F I C R

```

[illegible][illegible][illegible][illegible][illegible]

	S	L	T	G	M	T	-	S	L	I	R	P	W	F
	310		320		330		340		350		360			

[illegible][illegible]

```

      430      440      450      460      470      480
+1  C E S L E G S T P V R L A C T G C W H L F
+2  V R A L C K X A R L F A C A R G W D F O
+3  P L E H N V A L A L A L A L A L A L A L A
TGTGACAGCTTTCTCAAAACGACGCTTTCTCTCTCAACGCG
-1  ACACTCTCAACAGCAAGTTCCTCGCGACAGCAACCTCTCCGCAATCTCTCTGCA
-2  S L R L D M L V A T E R K H V P H I F S G L
-3  H S E L P F A C E T S T C R T V P V S L
-4  T L A Q G L A R R C C G C T B T V V E L

```

	420	500	510	520	530	540														
+1	K	T	E	Z	V	T	D	A	V	L	A	-	G	I	D	N	D	O	R	L
+2	R	L	E	N	-	O	T	L	C	-	R	E	V	L	T	O	T	N	V	S

Translation Results

D * R U D R R C A S V R Y * L O P T S L 5
 A A G T A C T A G A G O T G A C A G C T C T O T C T A C C T G A G O T A T T G A C T O G G A C C A A C T C T C
 T T G A C T A C T T C C A C T T C T C G A C A C A G A T C C A C T C C A T A A C T G A C C T T O G T C A C A G
 V B S T U 6 A T S A N P I F O S W P R Q
 S O L F L A L H Q A L T L Y O S P H U D R
 L S F L H C V R H - R R T N U V A - R T 6

550 560 570 580 590 600

C P K I L L N V P I S G I F S V A A P
A L K S L T G S L Q S V G S N L A O H L
T T T T A A A T T A C T G C C A T C C A T C C A T C T T T A G C C M A C T
A C G A A T T T T A C A T A G C C A C C A G G A G T T A T C A C C T A G G T A G A T A
R L I K S R O T G G I L P T S D T A A G
P R V S T G E L A H S G M O L L V P
A R F D A

[illegible][illegible]

730	740	750	760	770	780
					28
					29

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CROSS REFERENCES.

REST 202 29

Conclusions (continued)

* MERDSH appears to be what I had localized

The ATG here looks to be a great RoZak consensus

ACCATGG.

This is it

ORDER PRIMERS

LBRI #147 amino acid Sequence

MERDSDGNASPARTPSAGASPAQVSPAGTPPGRASPAQASPAQASPAQTPPGRASPAQASPAQTPPGRASPG
 RASPAQASPAQASPALASLSRSSSSGRSSSASASVTTSPTRVYLVRATPVGAVPIRSSPARSAPATRATRES
 PGTSLPKFTWREGQKQLPLIGCVLLLIALLVSLIILFQFWQGHGTGIRYKEQRESCPKHAVRCDGVVDCKLKS
 DELGCVRFDDWKSLLKIYSGSSHQWLPICSSNWNDSYSEKTCQQLGFESAHRRTTEVAHRDFANSFSILRYNS
 TIQESLHRSECPQRSYISLQCSHCHGLRAMTGRIVGGALASDSKWPQVSLHFGTTHICGTLIDAQWVLTA
 HCFVFTREKVLGKVGACTSNLHQLPEASIAETIINSNYTDEDDYDIALMRLSKPLTLSAHIHPACLPM
 HGQTFSLNETCWTGFGKTRETDDKTSPLFLEQVQNLIDFKKCNLDVYDSYLTPRMCMAGDLRGGRDSCQG
 DSGGPLVCEQNNRWYLAGVTSWGAGCGQRNKPVGVTYKVTEVLPWIYSKMESEVFRKS

Start is an NcoI site.

My predicted sequence diverges from the 5' end of BE280394 and a consensus splice acceptor (CAG).

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CR SS REFERENCES:

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BAYER CORPORATION

RB55202

SUBJECT

Oligo Order

EXHIBIT

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LBRI Oligos

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5

CoolEST 147- oligos to MY predicted genomic sequence

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RGL0054

Cloning primer 23-mer to BE280394 sequence

8

5' CTC AGA GAC CAT GGA GAG GGA CA 3'

9

10

RGL0055Sense Sequencing Primer 21-mer; bases 331-351 of
147.33.49.28contig2

11

12

5' CAA CCA GAG TGT ACC TTG TTA 3'

13

14

RGL0056antiense Sequencing Primer 20-mer; bases 441-460 of
147.33.49.28contig2

15

16

5' CAG GTG AAC TTG GGC AGG CT 3'

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Orcl

oligos to clone "Full length 147" shown on

Page 29

SIGNED BY

[Signature]

DATE

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CR SS REFERENCES:

BAYER CORPORATION

RB55202

SUBJECT PCR with RGL0054/0028

EXHIBIT

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Purpose: to test hypothesis for the new 5' ends

Received oligos

336178
R7181C08 (C08)
RGL0056
Rich Gedrich
284.66 ng (28 ng/100) OD 13.99
MW: 6180.0 u/gmole read 62.1
CAGGTGACCTTGGGACGCT

336178
R7181C08 (C08)
RGL0056
Rich Gedrich
456.23 ng (28 ng/100) OD 17.56
MW: 7732.8 u/gmole read 64.2
CTCAGCGACCATGCGAGGACCA

336178
R7181C07 (C07)
RGL0054
Rich Gedrich
286.11 ng (28 ng/100) OD 13.80
MW: 7732.8 u/gmole read 69.9
CTCAGCGACCATGCGAGGACCA

Resuspended RGL0054 to 200µl

added 250µl dH₂Odilute 1/24 to 50µl (25µl + 75µl dH₂O)

in 50µl PCR

ZLWS

do plasmid concn -/+ RT

in Advantage GC kit (Clontech)

Rows

1) - RT

2) + RT

Row

3µl (3)

2nd plasmid concn

10µl 5X Buffer

1µl Advantage GC polymerase

1µl 10mM dNTP

1µl RGL0054

1µl RGL0028

5µl GC mix

29µl dH₂O

50µl

1) Aliq + PM

2) Add concn

3) Cycle in PCT600

94°C 1' → 54°C 15" → 68°C 35" } 22 cycles
1 cycle } 1 cycle of

SIGNED BY

R. L. W. M.

DATE

WITNESSED AND UNDERSTOOD BY

Elizabeth C. Sullivan

DATE

CROSS REFERENCES:

BAYER CORPORATION

RB55202

41

SUBJECT



Run 5 gels on 1.2% Agarose gel (1X TAE)
M = 1 Kb Ladder (Gibco)

The Run was bad. A ~1.8 Kb Band was seen in the +RT Run only

Close to PCR

Run Remnants of the sample on a 1.2% Agarose gel

Cut out the Fragment

QIAquick Gel Extraction Kit Protocol

- using a microcentrifuge
- This protocol is designed to extract and purify DNA of 10 bp to 10 kb from standard or low melt agarose gels in TAE or TBE buffer.
- Notes:
- **NEW** The yellow color of Buffer OG indicates a pH of 7.5.
 - Add ethanol (95-100%) to Buffer PE before use (see bottle label for volume).
 - Isopropanol (100%) will be required.
 - A heating block or water bath at 50°C is required.
 - All centrifugation steps are carried out in a 10,000 x g (~13,000 rpm) in a conventional laboratory microcentrifuge.
 - 3M sodium acetate, pH 5.0, may be necessary.
1. Excise the DNA fragment from the agarose gel with a clean, sharp scalpel. Minimize the size of the gel slice by removing extra agarose.
 2. Weigh the gel slice in a colorless tube. Add 3 volumes of Buffer OG to 1 volume of gel (100 mg = 100 µl).
For example, add 300 µl of Buffer OG to each 100 mg of gel for >2% agarose gels, add 1 volume of Buffer OG. The maximum amount of gel slice per QIAquick column is 400 mg; for gel slices >400 mg use more than one QIAquick column.
 3. Incubate at 50°C for 10 min (or until the gel slice has completely dissolved). To help dissolve gel, mix by vortexing the tube every 2-3 min during the incubation.
IMPORTANT: Solubilize viscous completely. For >2% gels, increase incubation time.
 4. After the gel slice has dissolved completely, check that the color of the mixture is yellow (similar to Buffer OG without dissolved agarose). If the color of the mixture is orange or violet, add 10 µl of 3M sodium acetate, pH 5.0, and mix. The color of the mixture will turn to yellow.
The adsorption of DNA to QIAquick membrane is efficient only at pH 7.5. Buffer OG contains a pH indicator which is yellow at pH 7.5 and orange or violet at higher pH. Following entry determination of the optimal pH for DNA binding.
 5. Add 1 gel volume of isopropanol to the sample and mix.
For example, if the agarose gel slice is 100 mg, add 100 µl isopropanol. This step increases the yield of DNA fragments <500 bp and >4 kb. For DNA fragments between 500 bp and 4 kb, addition of isopropanol has no effect on yield. Do not centrifuge the sample at this stage.

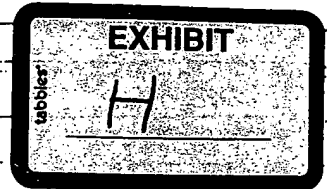
6. Place a QIAquick spin column in a provided 2-ml collection tube.
7. To bind DNA, apply the sample to the QIAquick column, and centrifuge for 1 min.
The maximum volume of the column reservoir is 800 µl. For sample volumes of more than 800 µl, apply load and spin again.
8. Discard flow-through and place QIAquick column back in the same collection tube.
Collection tubes are reused to reduce plastic consumption.
9. (Optional) Add 0.5 ml of Buffer OG to QIAquick column and centrifuge for 1 min.
This step will remove all traces of agarose. It is only required for direct sequencing, in vitro transcription or microinjection.
10. To wash, add 0.75 ml of Buffer PE to QIAquick column and centrifuge for 1 min.
Notes: If the DNA will be used for soft genome applications, such as library construction and direct sequencing, let the column stand 2-5 min after addition of Buffer PE, before centrifuging.
11. Discard the flow-through and centrifuge the QIAquick column for an additional 1 min at 10,000 x g (~13,000 rpm).
IMPORTANT: Residual alcohol from Buffer PE will not be completely removed unless the flow-through is discarded before the additional centrifugation.
12. Place QIAquick column into a clean 1.5-ml microcentrifuge tube.
13. To elute DNA, add 50 µl of Buffer EB (10 mM Tris-Cl, pH 8.5) or H₂O to the center of the QIAquick column and centrifuge for 1 min at maximum speed. Alternatively, for increased DNA concentration, add 30 µl elution buffer to the center of the QIAquick column, let stand for 1 min, and then centrifuge for 1 min.
IMPORTANT: Ensure that the elution buffer is dispensed directly onto the QIAquick membrane for complete elution of bound DNA. The average elution volume is 45 µl from 50 µl elution buffer volume, and 20 µl from 30 µl.
(Elution efficiency is dependent on pH. The maximum elution efficiency is achieved between pH 7.0 and 8.5. When using water, make sure that the pH is within this range, and store DNA at -20°C as DNA may degrade in the absence of a buffering agent. The purified DNA can also be stored in TE (10 mM Tris-Cl, 1 mM EDTA, pH 8.0), but the DNA may exhibit subsequent enzymatic reactions.)

SIGNED BY [Signature] DATE _____
WITNESSED AND UNDERSTOOD BY Elizabeth C. Sullivan DATE _____
CROSS REFERENCES:

BAYER CORPORATION

RB55202

SUBJECT



1

2

Eliel - 36 l dthd 147 0054/28 PER

3

4

TA cloning kit (Invitrogen)

5

6

Ligate Frg into pCR

7

Bsu

8

lab vector

9

lab 10x

10

4 l dthd

11

lab Lysate

12

3 l Fagat

13

Dad

14

16 °C o/p T_i = 550 pm

15

16

17

Transfer TDP 10

18

2nd Row / ~~pre~~ take

19

- 1 in 25'

20

- 42 °C 30"

21

- 1 in 2'

22

Add 400 ul 50x

23

37 °C 40"

24

25

plus 100 ul on LB + Amp Plate + Bluojet

26

27

37 °C o/p T_i

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SIGNED BY

A handwritten signature in cursive script, appearing to read "Elizabeth C. Sullivan".

DATE

WITNESSED AND UNDERST

OD BY

A handwritten signature in cursive script, appearing to read "Elizabeth C. Sullivan".

DATE

CR SS REFERENCE :

BAYER CORPORATION

RB5584L

7

SUBJECT LBRI #147

EXHIBIT

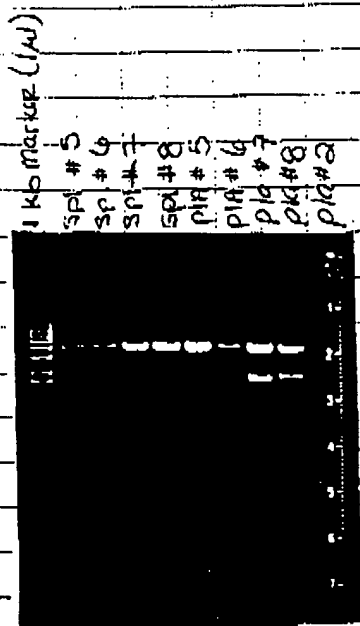
I

- ① miniprep of spl 5,6,7,8 : pla 5,6,7,8 : pla 2
 - * Followed same protocol from page 1 of this notebook.
 - * Eluted DNA in 50 μ l water

- ② Restriction enzyme digestion of pla 5,6,7,8, spl 5,6,7,8 : pla 2

Plasmid	3 μ l	Digest for 2 hrs @ 37°C
Eco RI	1 μ l	
Buffer 2	2 μ l	
H ₂ O	14 μ l	
	20 μ l	

- ③ RAN 1% Agarose Gel



* The miniprep from pla 2 gave negative results. Therefore, I did a new transformation using 1 μ l of a 1:200 dilution of my original pla 2 miniprep. Transformed in 30 \times of DH5 α max efficiency.

- 1 \times dilution + 30 \times cells
- incubate on ice for 5-10 minutes
- Add 100 \times SOC
- Plate 100 \times on Amp plates
- Incubate @ 37°C overnight

SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERSTOOD BY

Dr. James A. Delver

DATE

BAYER CORPORATION

SUBJECT LARI #147

① Miniprep of #1475428 (LARI)
 Rich Gedrich gave me 4 clones from spleen
 and 4 clones from placenta

② I followed the following protocol using 1.5 ml culture

QIAprep Spin Miniprep Kit Protocol

using a microcentrifuge

This protocol is designed for purification of up to 20 µg of high-copy plasmid DNA from 1-5 ml overnight cultures of *E. coli* in LB [Luria-Bertani] medium. For purification of low-copy plasmids and cosmids, large plasmids (>10 kb), and DNA prepared using other methods, refer to the recommendations on page 31.

Please read Important Notes for QIAprep Procedures on pages 14-15 before starting.

Procedure

1. Resuspend pelleted bacterial cells in 250 µl of Buffer P1 and transfer to a microfuge tube.
 Ensure that RNase A has been added to Buffer P1. No cell clumps should be visible after resuspension of the pellet.
2. Add 250 µl of Buffer P2 and gently invert the tube 4-6 times to mix.
 Mix gently by inverting the tube. Do not vortex, as this will result in shearing of genomic DNA. If necessary, continue inverting the tube until the solution becomes viscous and slightly clear. Do not allow the lysis reaction to proceed for more than 5 min.
3. Add 350 µl of Buffer N3 and invert the tube immediately but gently 4-6 times.
 To avoid localized precipitation, mix the solution gently but thoroughly, immediately after addition of Buffer N3. The solution should become cloudy.
4. Centrifuge for 10 min.
 A compact white pellet will form.
 During centrifugation, place a QIAprep spin column in a 2-ml collection tube.
5. Apply the supernatant from step 4 to the QIAprep column by decanting or pipetting.
6. Centrifuge 30-60 sec. Discard the flow-through.
7. [Optional]: Wash QIAprep spin column by adding 0.5 ml of Buffer PB and centrifuging 30-60 sec. Discard the flow-through.
 This step is necessary to remove trace nuclease activity when using endA⁻ strains such as the JM series, HB101 and its derivatives, or any wild-type strain, which have high levels of nuclease activity or high carbohydrate content. Host strains such as XL-1 Blue and DH5α™ do not require this additional wash step.
8. Wash QIAprep spin column by adding 0.75 ml of Buffer PE and centrifuging 30-60 sec.

9. Discard the flow-through, and centrifuge for an additional 1 min to remove residual wash buffer.

! IMPORTANT: Residual wash buffer will not be completely removed unless the flow-through is discarded before this additional centrifugation. Residual ethanol from Buffer PE may inhibit subsequent enzymatic reactions.

10. Place QIAprep column in a clean 1.5-ml microfuge tube. To elute DNA, add 50 µl of Buffer EB (10 mM Tris-Cl, pH 8.5) or H₂O to the center of each QIAprep column, let stand for 1 min, and centrifuge for 1 min.

QIAprep Spin Miniprep Kit Protocol

using 5-ml collection tubes

The QIAprep Spin Miniprep procedure can be performed using 5-ml centrifuge tubes (e.g., Greiner, Cat. No. 115101 or 115261) as collection tubes to decrease handling. The standard protocol on pages 18-19 should be followed with the following modifications:

- Step 4: Place QIAprep spin column in a 5-ml centrifuge tube instead of a 2-ml collection tube.
- Step 6: Centrifuge at 3000 x g for 1 min using a suitable rotor (e.g., Beckman® GS-6KR centrifuge at ~6000 rpm). (The flow-through does not need to be discarded).
- Steps 7 & 8: For washing steps, centrifugation should be performed at 3000 x g for 1 min. (The flow-through does not need to be discarded).
- Step 9: Transfer QIAprep column to a microfuge tube. Microcentrifuge at maximum speed for 1 min. Continue with step 10 of the protocol.

- * Pellet cells at 10,000 rpm for 2 minutes
- * ~~Elute~~ Elute DNA in 50 µl's water instead of Buffer EB
- next time I do a miniprep
- * Store excess bacterial culture in 4°C

SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERSTOOD BY

Angela A. Decker

DATE

BAYER CORPORATION

RB5T846

EXHIBIT

SUBJECT LBRI #147

tabbies

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Lisa Parkyn

03:35 PM

To: Janice Jackson/WESTH/PH/US/BAYER@BAYER-US-NOTES, Gwenda
Ligon/WESTH/PH/US/BAYER@BAYER-US-NOTES
cc: David Eustice/WESTH/PH/US/BAYER@BAYER-US-NOTES

Subject: LBRI Sequencing

Hi Jan and Gwen,

These are for the LBRI #147 program.

The vector is pCRII. The host was Top10 and the preps were done with the Qiagen kit.

Thanks,

Lisa

1. RG_spi5_M13F
2. RG_spi5_029
3. RG_spi5_032
4. RG_spi5_033
5. RG_spi5_034
6. RG_spi5_047
7. RG_spi5_052
8. RG_spi5_M13R
9. RG_spi6_M13F
10. RG_spi6_029
11. RG_spi6_032
12. RG_spi6_033
13. RG_spi6_034
14. RG_spi6_047
15. RG_spi6_052
16. RG_spi6_M13R
17. RG_pla6_M13F
18. RG_pla6_029
19. RG_pla6_032
20. RG_pla6_033
21. RG_pla6_034
22. RG_pla6_047
23. RG_pla6_052
24. RG_pla6_M13R
25. RG_pla7_M13F
26. RG_pla7_029
27. RG_pla7_032
28. RG_pla7_033
29. RG_pla7_034
30. RG_pla7_047
31. RG_pla7_052
32. RG_pla7_M13R
33. RG_pla8_M13F
34. RG_pla8_029
35. RG_pla8_032

spi5, spi6, pla6, pla2

3x plasmid
2x primer
7x H₂O

pla7, pla8

1x plasmid
2x primer
9x H₂O

Primers

M13F, RG029, 032, 033,
034, 047, 052, M13R

36. RG_pla8_033
37. RG_pla8_034
38. RG_pla8_047
39. RG_pla8_052
40. RG_pla3_M13R
41. RG_pla3_M13F
42. RG_pla3_029
43. RG_pla3_032
44. RG_pla3_033
45. RG_pla3_034
46. RG_pla3_047
47. RG_pla3_052
48. RG_pla3_M13R

SIGNED BY

Lisa Parkyn

DATE

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Dargues A - DeHue

DATE

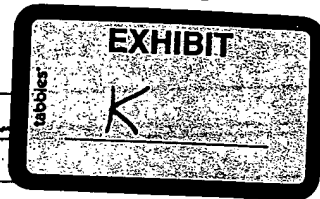
CROSS REFERENCES:

BAYER CORPORATION

RB55846

15

SUBJECT LBRI #147



① LifeTools Clustal W results comparing coolEST147_147spl5_2
the 147 consensus sequence (DNA sequence) (per vector)

Confidential - Property of Incyte Genomics, Inc. LifeTools Version 3.1 SeqServer

coolEST147_147spl5_2
coolEST147_147FLconsensus_2

CLUSTAL W (1.7) Multiple Sequence Alignments

Sequence format is Pearson
Sequence 1: coolEST147_147spl5_2 1767 bp
Sequence 2: coolEST147_147FLconsensus_2 1748 bp
Start of Pairwise alignments
Aligning...
Sequences (1:2) Aligned. Score: 100
Start of Multiple Alignment
There are 1 groups
Aligning...
Group 1: Sequences: 2 Score: 33212
Alignment Score 13534
CLUSTAL-Alignment file created (baaa03540.aln)
CLUSTAL W (1.7) multiple sequence alignment

```

coolEST147_147spl5_2      GAATTCGGCTCAGAGACCATGGAGAGGCACAGCCACGGGAATGCATCTCC
coolEST147_147FLconsensus_2 -----CTCAGAGACCATGGAGAGGCACAGCCACGGGAATGCATCTCC
*****

coolEST147_147spl5_2      AGCAAGAACACCTTCAGCTGGAGCATCTCCAGCCAGGCATCTCCAGCTG
coolEST147_147FLconsensus_2 AGCAAGAACACCTTCAGCTGGAGCATCTCCAGCCAGGCATCTCCAGCTG
*****

coolEST147_147spl5_2      GGACACCTCCAGGCCGGGCATCTCCAGCCAGGCATCTCCAGCCAGGCA
coolEST147_147FLconsensus_2 GGACACCTCCAGGCCGGGCATCTCCAGCCAGGCATCTCCAGCCAGGCA
*****

coolEST147_147spl5_2      TCTCCAGCTGGACACCTCCGGCCGGGCATCTCCAGCCAGGCATCTCC
coolEST147_147FLconsensus_2 TCTCCAGCTGGACACCTCCGGCCGGGCATCTCCAGCCAGGCATCTCC
*****

coolEST147_147spl5_2      AGCTGGTACACCTCCAGGCCGGGCATCTCCAGCCAGGCATCTCCAGCCC
coolEST147_147FLconsensus_2 AGCTGGTACACCTCCAGGCCGGGCATCTCCAGGCCGGGCATCTCCAGCCC
*****

coolEST147_147spl5_2      AGGCATCTCCAGCCCGGGCATCTCCGGCTCTGGCATCACTTTCCAGGTCC
coolEST147_147FLconsensus_2 AGGCATCTCCAGCCCGGGCATCTCCGGCTCTGGCATCACTTTCCAGGTCC
*****

coolEST147_147spl5_2      TCATCCGGCAGGTGATCATCCGCCAGGTGAGCTCGGTGACAACTCCCC
coolEST147_147FLconsensus_2 TCATCCGGCAGGTGATCATCCGCCAGGTGAGCTCGGTGACAACTCCCC
*****

coolEST147_147spl5_2      AACCAGAGTGATCTTGTAGAGCAACACCAAGTGGGGGCTGTACCCATCC
coolEST147_147FLconsensus_2 AACCAGAGTGATCTTGTAGAGCAACACCAAGTGGGGGCTGTACCCATCC
*****

coolEST147_147spl5_2      GATCATCTCTGCCAGGTGAGCACCAGCAACAGGGCCACAGGGAGAGC
coolEST147_147FLconsensus_2 GATCATCTCTGCCAGGTGAGCACCAGCAACAGGGCCACAGGGAGAGC
*****
    
```

*Continued on
pg. 16

SIGNED BY Olga Parkyn

DATE

WITNESSED AND UNDERSTOOD BY Dr. Wayne J. Diller

DATE

----- REFERENCES -----

SUBJECT LBRI #147

1	LifeTools Clustal W... (DNA Sequence)
2	
3	coolEST147_147spl5_2 CCAGGTACGAGCCTGCCAAGTTCACCTGGCGGAGGGCCAGAAGCAGCT
4	coolEST147_147FLconsensus_2 CCAGGTACGAGCCTGCCAAGTTCACCTGGCGGAGGGCCAGAAGCAGCT
5	coolEST147_147spl5_2 ACCGCTCATCGGGTGCGTCTCTCTCTGCGCTGGTGGTTTCGCTCA
6	coolEST147_147FLconsensus_2 ACCGCTCATCGGGTGCGTCTCTCTCTGCGCTGGTGGTTTCGCTCA
7	coolEST147_147spl5_2 TCATCTCTTCCAGTTCTGGCAGGGCCACACAGGGATCAGGTACAAGGAG
8	coolEST147_147FLconsensus_2 TCATCTCTTCCAGTTCTGGCAGGGCCACACAGGGATCAGGTACAAGGAG
9	coolEST147_147spl5_2 CAGACGGAGAGCTGTCCCAAGCACGCTGTTCGCTGTGACGGGGTGGTGA
10	coolEST147_147FLconsensus_2 CAGACGGAGAGCTGTCCCAAGCACGCTGTTCGCTGTGACGGGGTGGTGA
11	coolEST147_147spl5_2 CTGCAAGCTGAAGAGTGACGAGCTGGGCTGCGTGAGGTTTGACTGGGACA
12	coolEST147_147FLconsensus_2 CTGCAAGCTGAAGAGTGACGAGCTGGGCTGCGTGAGGTTTGACTGGGACA
13	coolEST147_147spl5_2 AGTCTCTGCTTAAATCTACTCTGGGTCTCCCATCAGTGGCTTCCCATC
14	coolEST147_147FLconsensus_2 AGTCTCTGCTTAAATCTACTCTGGGTCTCCCATCAGTGGCTTCCCATC
15	coolEST147_147spl5_2 TGTAGCAGCAACTGGAATGACTCCTACTCAGAGAAGACCTGCCAGCAGCT
16	coolEST147_147FLconsensus_2 TGTAGCAGCAACTGGAATGACTCCTACTCAGAGAAGACCTGCCAGCAGCT
17	coolEST147_147spl5_2 GGGTTTCGAGAGTGCTCACCGGACAACCGAGCTTGCCACAGGGATTGTG
18	coolEST147_147FLconsensus_2 GGGTTTCGAGAGTGCTCACCGGACAACCGAGCTTGCCACAGGGATTGTG
19	coolEST147_147spl5_2 CCAACAGCTTCTCAATCTTGAGATACAACTCCACCATCCAGGAAGCCTC
20	coolEST147_147FLconsensus_2 CCAACAGCTTCTCAATCTTGAGATACAACTCCACCATCCAGGAAGCCTC
21	coolEST147_147spl5_2 CACAGGTCTGAATGCCCTTCCAGCGGTATATCTCCCTCCAGTGTCCCA
22	coolEST147_147FLconsensus_2 CACAGGTCTGAATGCCCTTCCAGCGGTATATCTCCCTCCAGTGTCCCA
23	coolEST147_147spl5_2 CTGCGGACTGAGGGCCATGACCGCGGATCTGGGAGGGCGCTGGCTT
24	coolEST147_147FLconsensus_2 CTGCGGACTGAGGGCCATGACCGCGGATCTGGGAGGGCGCTGGCTT
25	coolEST147_147spl5_2 CGGATAGCAAGTGGCTTGGCAAGTGAGTCTGCACTTGGCACCACCCAC
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29	coolEST147_147spl5_2 CTGCTTCTCTGACCCCGGAGAGGTCTGGAGGGCTGGAAGGTGTACG
30	coolEST147_147FLconsensus_2 CTGCTTCTCTGACCCCGGAGAGGTCTGGAGGGCTGGAAGGTGTACG
31	coolEST147_147spl5_2 CGGGCACCAGCAACCTGCACCACTTGCCTGAGGCAGCCTCCATTGCCGAG
32	coolEST147_147FLconsensus_2 CGGGCACCAGCAACCTGCACCACTTGCCTGAGGCAGCCTCCATTGCCGAG
33	coolEST147_147spl5_2 ATCATCATCAACAGCAATTACACCGATGAGGAGGACGACTATGACATGC
34	coolEST147_147FLconsensus_2 ATCATCATCAACAGCAATTACACCGATGAGGAGGACGACTATGACATGC
	CCTCATGCGGCTGTCCAAAGCCCTGACCTGTCCGCTCAGTCCACCCCTG
	CCTCATGCGGCTGTCCAAAGCCCTGACCTGTCCGCTCAGTCCACCCCTG
	CTTGCCCTCCCATGCAATGGACAGACCTTACGCTCAATGAGACCTGCTGG
	CTTGCCCTCCCATGCAATGGACAGACCTTACGCTCAATGAGACCTGCTGG

continued on
pg. 17.

SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERST OD BY

Dagmar A. Diller

DATE

ADVICE RECEIVED.

BAYER CORPORATION

RB55846 17

SUBJECT LBRI #147

Lifetools Clustal w (DNA Sequence)

coolEST147_147sp15_2	ATCACAGGCTTTGGCAAGACCAGGAGACAGATGACAAGACATCCCCCTT	1
coolEST147_147FLconsensus_2	ATCACAGGCTTTGGCAAGACCAGGAGACAGATGACAAGACATCCCCCTT	2
coolEST147_147sp15_2	CCTCCGGGAGGTGCAGGTCAATCTCATCGACTTCAAGAAATGCAATGACT	3
coolEST147_147FLconsensus_2	CCTCCGGGAGGTGCAGGTCAATCTCATCGACTTCAAGAAATGCAATGACT	4
coolEST147_147sp15_2	ACTTGGTCTATGACAGTTACCTTACCCCAAGGATGATGTGTCTGGGGAC	5
coolEST147_147FLconsensus_2	ACTTGGTCTATGACAGTTACCTTACCCCAAGGATGATGTGTCTGGGGAC	6
coolEST147_147sp15_2	CTTCGTGGGGCCAGAGACTCCTGCCAGGGAGACAGCGGGGGCCCTTTGT	7
coolEST147_147FLconsensus_2	CTTCGTGGGGCCAGAGACTCCTGCCAGGGAGACAGCGGGGGCCCTTTGT	8
coolEST147_147sp15_2	CTGTGAGCAGAAACCCGCTGGTACCTGGCAGGTGTCAACAGCTGGGGCA	9
coolEST147_147FLconsensus_2	CTGTGAGCAGAAACCCGCTGGTACCTGGCAGGTGTCAACAGCTGGGGCA	10
coolEST147_147sp15_2	CAGGCTGTGGCCAGAGAAACAAACCTGGTGTGTACACAAAGTGACAGAA	11
coolEST147_147FLconsensus_2	CAGGCTGTGGCCAGAGAAACAAACCTGGTGTGTACACAAAGTGACAGAA	12
coolEST147_147sp15_2	GTTCCTCCCTGGATTACACCAAGATGGAGAGCGAGGTCCGATTAGAAA	13
coolEST147_147FLconsensus_2	GTTCCTCCCTGGATTACACCAAGATGGAGAGCGAGGTCCGATTAGAAA	14
coolEST147_147sp15_2	ATCCTAACCAAGCTGGCTGCTGCTCTGCACAGCACCAGGCTGCTGTGACTC	15
coolEST147_147FLconsensus_2	ATCCTAACCAAGCTGGCTGCTGCTCTGCACAGCACCAGGCTGCTGTGACTC	16
coolEST147_147sp15_2	GAGAAAAAGCCGAATTC	17
coolEST147_147FLconsensus_2	GAGAAA	18

Protein Sequence

Sequence format is Pearson
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 Sequence 2: coolEST147_147FLconsensus_3 562 aa
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 Aligning...
 Sequences (1:2) Aligned. Score: 100
 Start of Multiple Alignment
 There are 1 groups
 Aligning...
 Group 1: Sequences: 2 Score: 7726
 Alignment Score 3604
 CLUSTAL-Alignment file created [baaa03SM3.aln]
 CLUSTAL W (1.7) multiple sequence alignment

coolEST147_147sp15+1_1	MERDSHGNASPARTPSAGASPAQASPAQTTPGRASPAQASPAQASPAQTTP	19
coolEST147_147FLconsensus_3	MERDSHGNASPARTPSAGASPAQASPAQTTPGRASPAQASPAQASPAQTTP	20
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coolEST147_147FLconsensus_3	PGRASPAQASPAQTTPGRASPGRASPAQASPARASPALASLSRSSSSGRSS	22
coolEST147_147sp15+1_1	SARSASVTTSPTRVYLVRATPVCAVPIRSSPARSAPATRATRESPTSLP	23
coolEST147_147FLconsensus_3	SARSASVTTSPTRVYLVRATPVCAVPIRSSPARSAPATRATRESPTSLP	24

*continued on
 pg. 18

SIGNED BY

Olga Paulsen

DATE

WITNESSED AND UNDERSTOOD BY

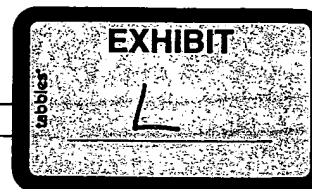
Darguise A. DeHeer

DATE

BAYER CORPORATION

RB55846 17

SUBJECT LBRI #147



Lifetools Clustal w (DNA Sequence)

coolEST147_147spl5_2	ATCACAGGCTTTGGCAAGACCAGGAGACAGATGACAAGACATCCCCCTT	1
coolEST147_147FLconsensus_2	ATCACAGGCTTTGGCAAGACCAGGAGACAGATGACAAGACATCCCCCTT	2
coolEST147_147spl5_2	CCTCCGGGAGGTGCAGGTCAATCTCATCGACTTCAAGAAATGCAATGACT	3
coolEST147_147FLconsensus_2	CCTCCGGGAGGTGCAGGTCAATCTCATCGACTTCAAGAAATGCAATGACT	4
coolEST147_147spl5_2	ACTTGGTCTATGACAGTTACCTTACCCCAAGGATGATGTGTCTGGGGAC	5
coolEST147_147FLconsensus_2	ACTTGGTCTATGACAGTTACCTTACCCCAAGGATGATGTGTCTGGGGAC	6
coolEST147_147spl5_2	CTTCGTGGGGCCAGAGACTCTCTGCCAGGAGACACCGGGGGCCTCTTGT	7
coolEST147_147FLconsensus_2	CTTCGTGGGGCCAGAGACTCTCTGCCAGGAGACACCGGGGGCCTCTTGT	8
coolEST147_147spl5_2	CTGTGAGCAGAAACACCGCTGGTACCTGGCAGGTGTACCCAGCTGGGCA	9
coolEST147_147FLconsensus_2	CTGTGAGCAGAAACACCGCTGGTACCTGGCAGGTGTACCCAGCTGGGCA	10
coolEST147_147spl5_2	CAGGCTGTGGCCAGAGAAACAAACCTGGTGTGTACACCAAGTGACAGAA	11
coolEST147_147FLconsensus_2	CAGGCTGTGGCCAGAGAAACAAACCTGGTGTGTACACCAAGTGACAGAA	12
coolEST147_147spl5_2	GTTCCTCCCTGGATTTACAGCAAGATGGAGAGCGAGGTCCGATTGAGAA	13
coolEST147_147FLconsensus_2	GTTCCTCCCTGGATTTACAGCAAGATGGAGAGCGAGGTCCGATTGAGAA	14
coolEST147_147spl5_2	ATCCTAACCCAGCTGGCCTGCTGCTCTGCACAGCACCGGCTGCTGTGACTC	15
coolEST147_147FLconsensus_2	ATCCTAACCCAGCTGGCCTGCTGCTCTGCACAGCACCGGCTGCTGTGACTC	16
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coolEST147_147FLconsensus_2	GAGAAA	18

Protein Sequence

Sequence format is Pearson
 Sequence 1: coolEST147_147spl5-1_1 562 aa
 Sequence 2: coolEST147_147FLconsensus_3 562 aa
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 Aligning...
 Sequences (1:2) Aligned. Score: 100
 Start of Multiple Alignment
 There are 1 groups
 Aligning...
 Group 1: Sequences: 2 Score: 7726
 Alignment Score 1604
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 CLUSTAL W (1.7) multiple sequence alignment

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coolEST147_147spl5-1_1	PGRASPAQASPAQTPPGRASPAQASPAQASPARASPALASLSRSSSGRSS	21
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coolEST147_147FLconsensus_3	SARSASVTTSPTRVYLVRATPVGAVPIRSSPARSAPATRATRESPTSLP	24

*continued on
 pg. 18

SIGNED BY

Don Paulsen

DATE

WITNESSED AND UNDERSTOOD BY

Duggan A. DeHe

DATE

SUBJECT LBRI #147

1	<u>Protein sequence (Life tools)</u>	
2		
3	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	KFTWREGQKQLPLIGCVLLIALVVSLLILFQFWQGHGTGIRYKEQRESCP KFTWREGQKQLPLIGCVLLIALVVSLLILFQFWQGHGTGIRYKEQRESCP *****
4		
5	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	KHAVRCDCGVVDCCKLSDELQCVRFWDKSLKLYSGSSHQWLPICSSNNW KHAVRCDCGVVDCCKLSDELQCVRFWDKSLKLYSGSSHQWLPICSSNNW *****
6		
7	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	DSYSEKTCQQLGFESAHRTTEVAHRDFANSFSLRYNSTIQESLHRSECP DSYSEKTCQQLGFESAHRTTEVAHRDFANSFSLRYNSTIQESLHRSECP *****
8		
9	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	SQRYISLQCSHCGLRAMTGRIVGGALASDSKWPQVSLHFGTTHICGGTL SQRYISLQCSHCGLRAMTGRIVGGALASDSKWPQVSLHFGTTHICGGTL *****
10		
11	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	IDAQWVLTAAHCFFVTREKVLGKWKVYAGTSLNLHQLPEAASIAEIIINSN IDAQWVLTAAHCFFVTREKVLGKWKVYAGTSLNLHQLPEAASIAEIIINSN *****
12		
13	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	YTDEEDDYDIALMRLSKPLTSLAHIHAPCLPMHGQTFSLNETCWTGFGK YTDEEDDYDIALMRLSKPLTSLAHIHAPCLPMHGQTFSLNETCWTGFGK *****
14		
15	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	TRETDDKTSPPFLREVQVNLIDFKKCNLYLVYDSYLTPRMMCAGDLRGGRD TRETDDKTSPPFLREVQVNLIDFKKCNLYLVYDSYLTPRMMCAGDLRGGRD *****
16		
17	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	SCQGDGSGGPLVCEQNNRWYLAGVTSWGTGCGQRNKPQVYTKVTEVLPWIY SCQGDGSGGPLVCEQNNRWYLAGVTSWGTGCGQRNKPQVYTKVTEVLPWIY *****
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SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERSTOOD BY

Dargines A. Adler

DATE

CROSS REFERENCES: